

KICK OFF MEETING

FRIDAY MARCH 23rd, 2018  
PARIS

International Association for Responsible  
Research and Innovation in Genome Editing  
(ARRIGE)

# Setting the stage, where do we stand today with CRISPR technology

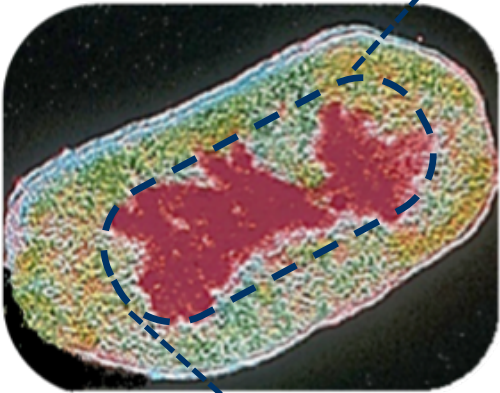
**Francisco J.M. Mojica**



Universitat d'Alacant  
Universidad de Alicante

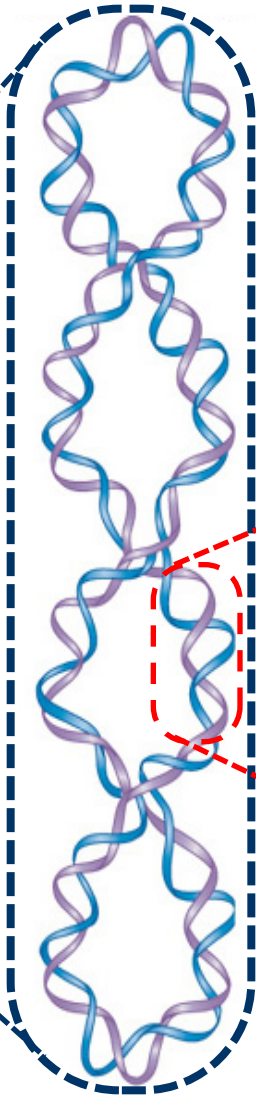
# Plain CRISPR

Prokaryotes



The MacGraw-Hill Companies.  
Microbiology, by Precott, Harley & Klein. 7th Ed.

Genome



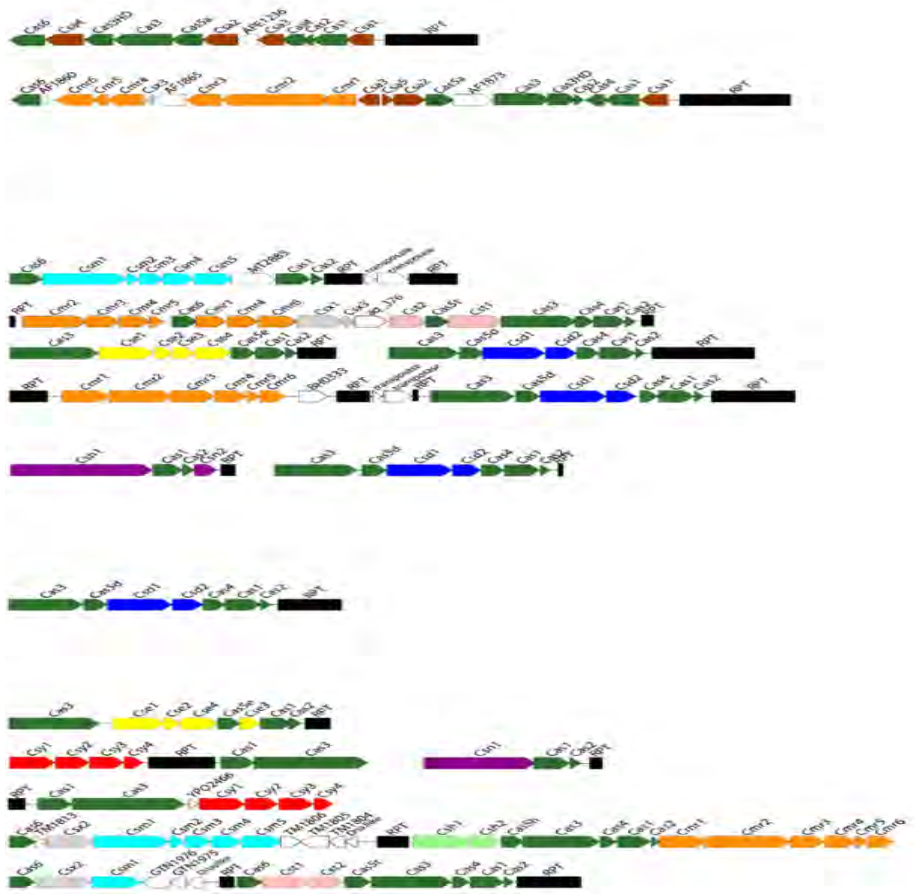
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GGCGGGCGGTTACAGACGAACCCTAGTTGGGTTGAAGCGACAAGACTCGCGACGAAGCCGAGT  
CGAAACGCCGCGTTACAGACGAACCCTAGTTGGGTTGAAGCCTCTTTATCCCTCCTGCCCGAA  
TGCTACGAATATCGTTACAGACGAACCCTAGTTGGGTTGAAGCGAACCCACTGGTGAAGAAA  
AAGTTGTAGAGACCCTAGTTACAGACGAATCCCTAGTTGGGTTGAAGCAGCACAATCAAGTCT  
GGTTACATGGCGACAGGATGGGTTACAGACGAACCCTAGTTGGGTTGAAGCTTCCACAACGTC  
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```

# Spacers



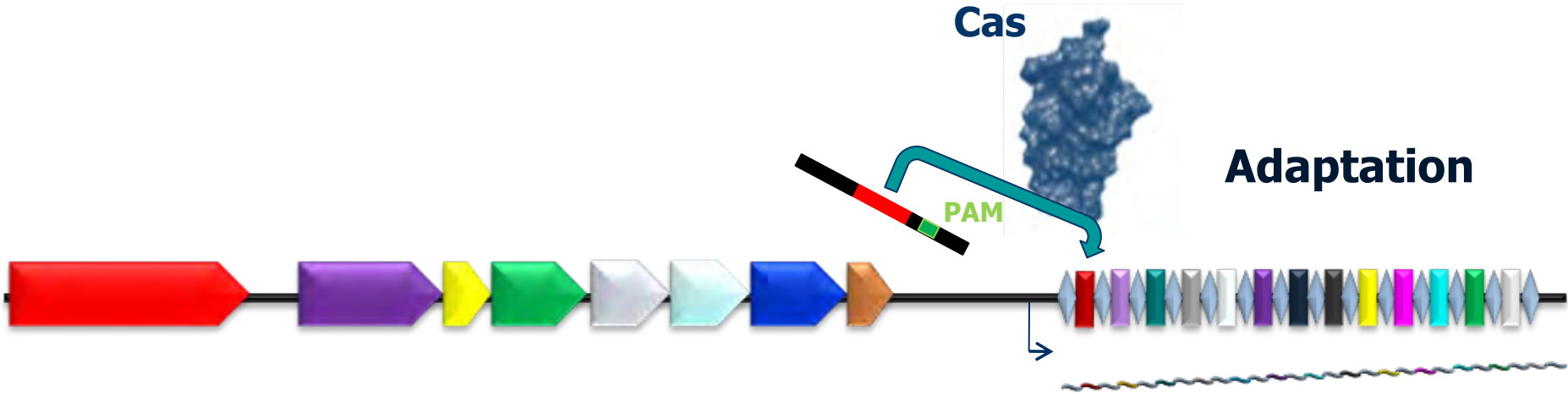
*cas* (*CRISPR associated*)

◆ **C R I S P R**  
 l e n h a e  
 u g t o l p  
 s u e r i e  
 t l r t n a  
 e a s d t  
 r r p r  
 e l a o  
 d y c m  
 e i  
 d c



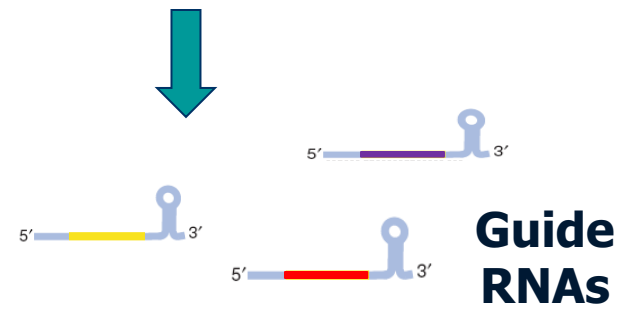
Modified from Haft et al. PLoS Comput. Biol. 2005

# **The CRISPR Mechanism**

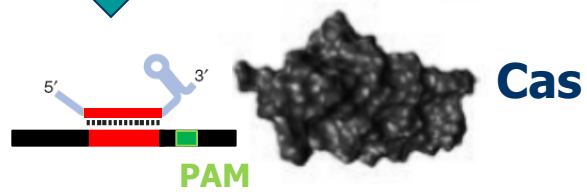


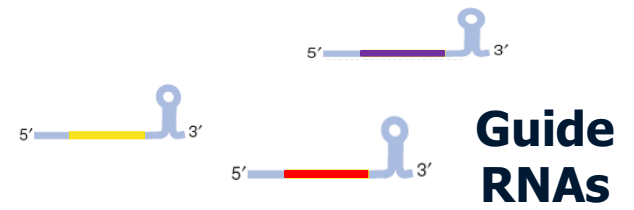
**Expression**

Cas

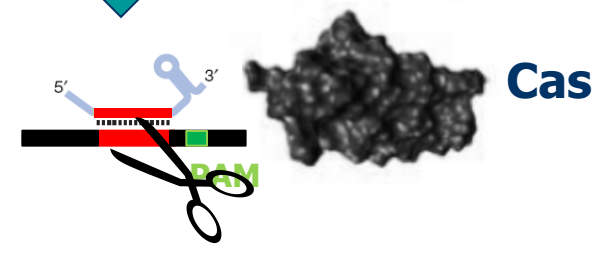


**Interference**





**Interference**



# Programmable DNA cleavage

PNAS

## Cas9–crRNA ribonucleoprotein complex mediates specific DNA cleavage for adaptive immunity in bacteria

Giedrius Gasiunas<sup>a</sup>, Rodolphe Barrangou<sup>b</sup>, Philippe Horvath<sup>c</sup>, and Virginijus Siksnys<sup>a,1</sup>

<sup>a</sup>Institute of Biotechnology, Vilnius University, LT-02241 Vilnius, Lithuania; <sup>b</sup>DuPont Nutrition and Health, Madison, WI 53716; and <sup>c</sup>DuPont Nutrition and Health, F-86220 Dange-Saint-Romain, France

Edited by Arthur Landy, Brown University, Providence, RI, and approved August 1, 2012 (received for review May 21, 2012)



R. Barrangou



P. Horvath



V. Siksnys



## A Programmable Dual-RNA–Guided DNA Endonuclease in Adaptive Bacterial Immunity

Martin Jinek *et al.*

*Science* 337, 816 (2012);

DOI: 10.1126/science.1225829

*"... to exploit the system for RNA-programmable genome editing"*



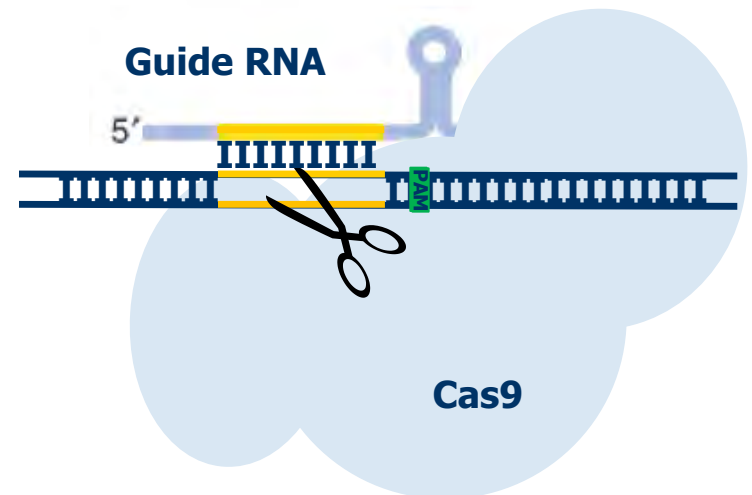
E. Charpentier



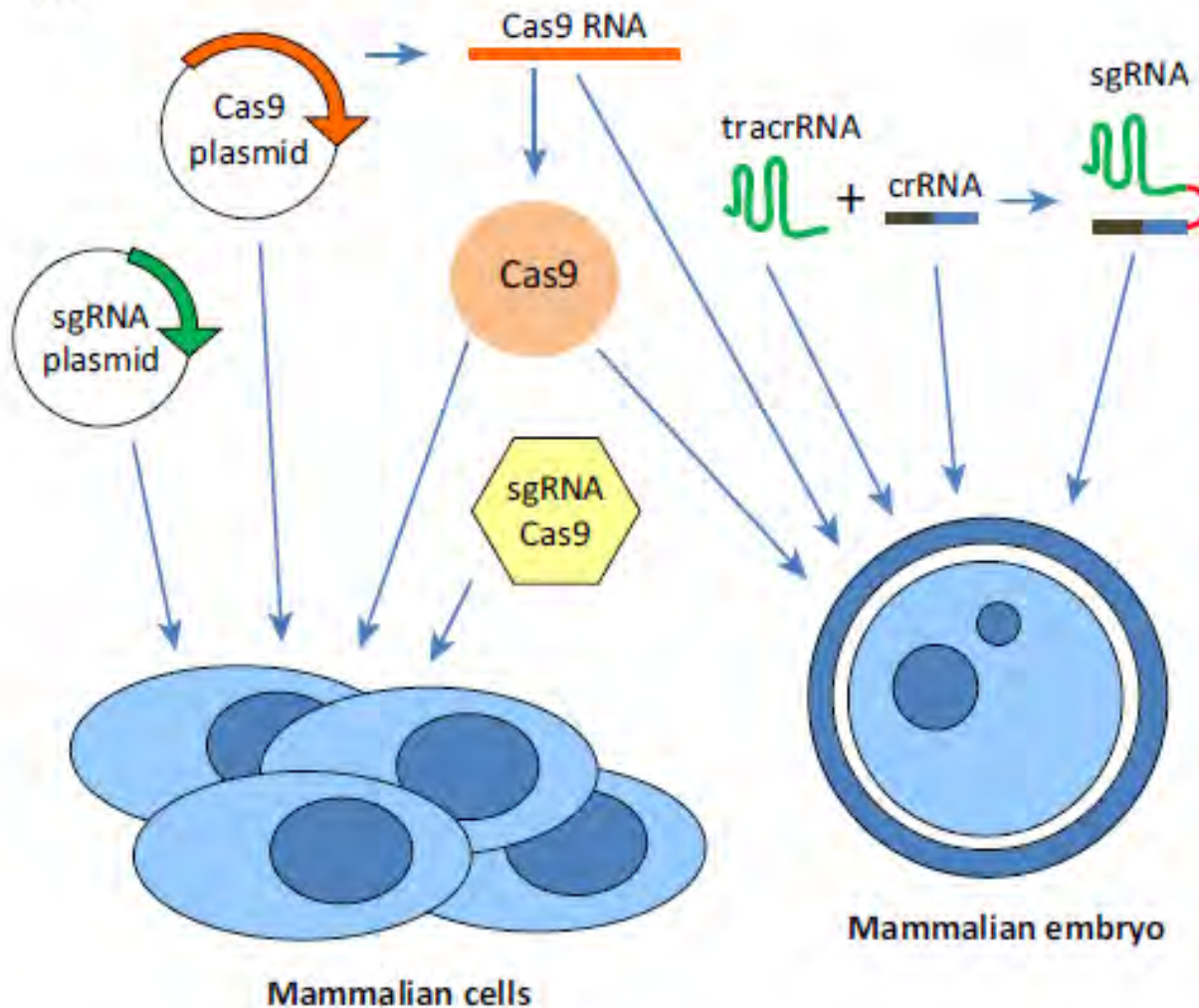
J. Doudna



DNA



# CRISPR-Cas9 delivery



Trends in Microbiology

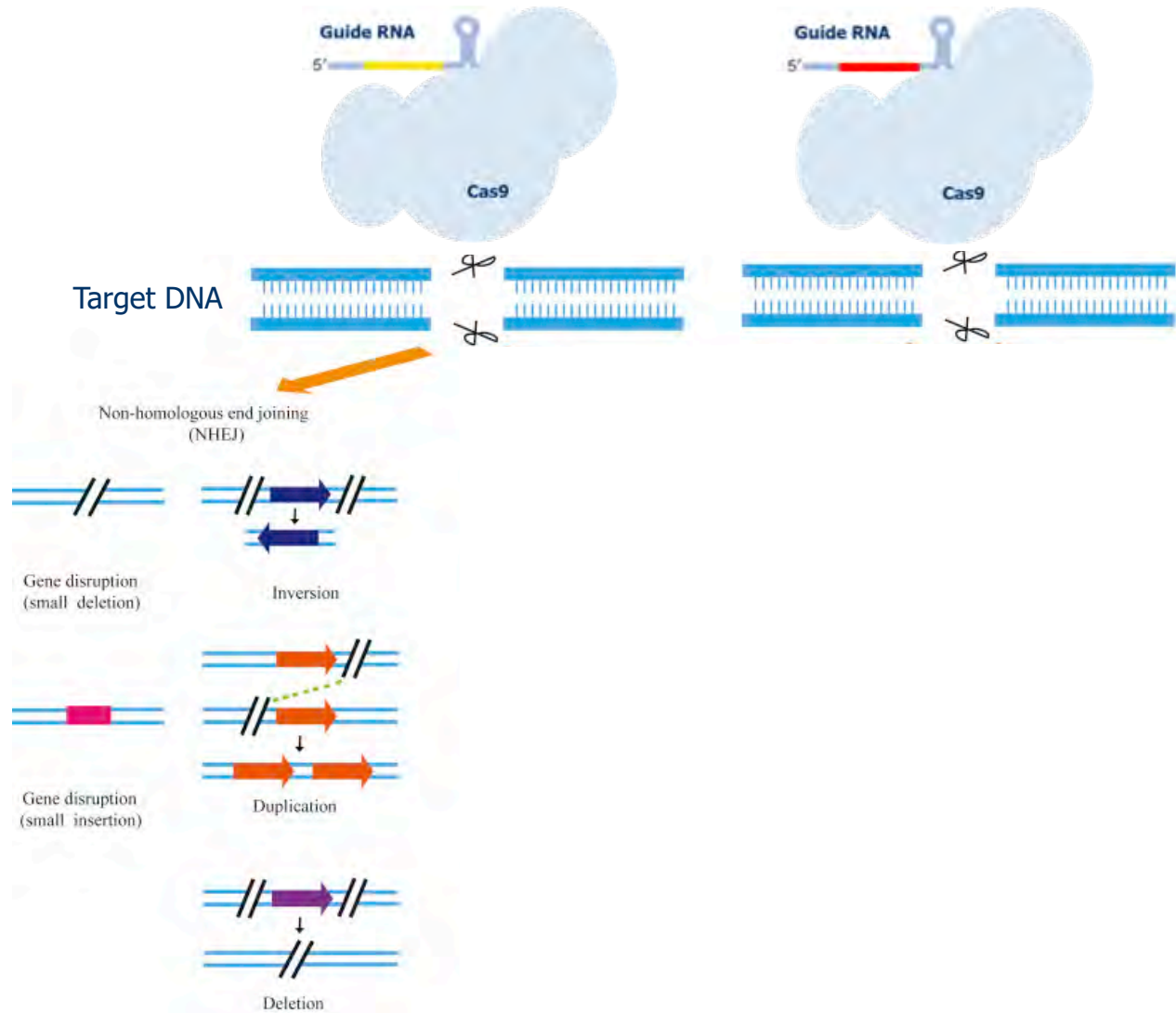
## Review

On the Origin of CRISPR-Cas Technology: From Prokaryotes to Mammals

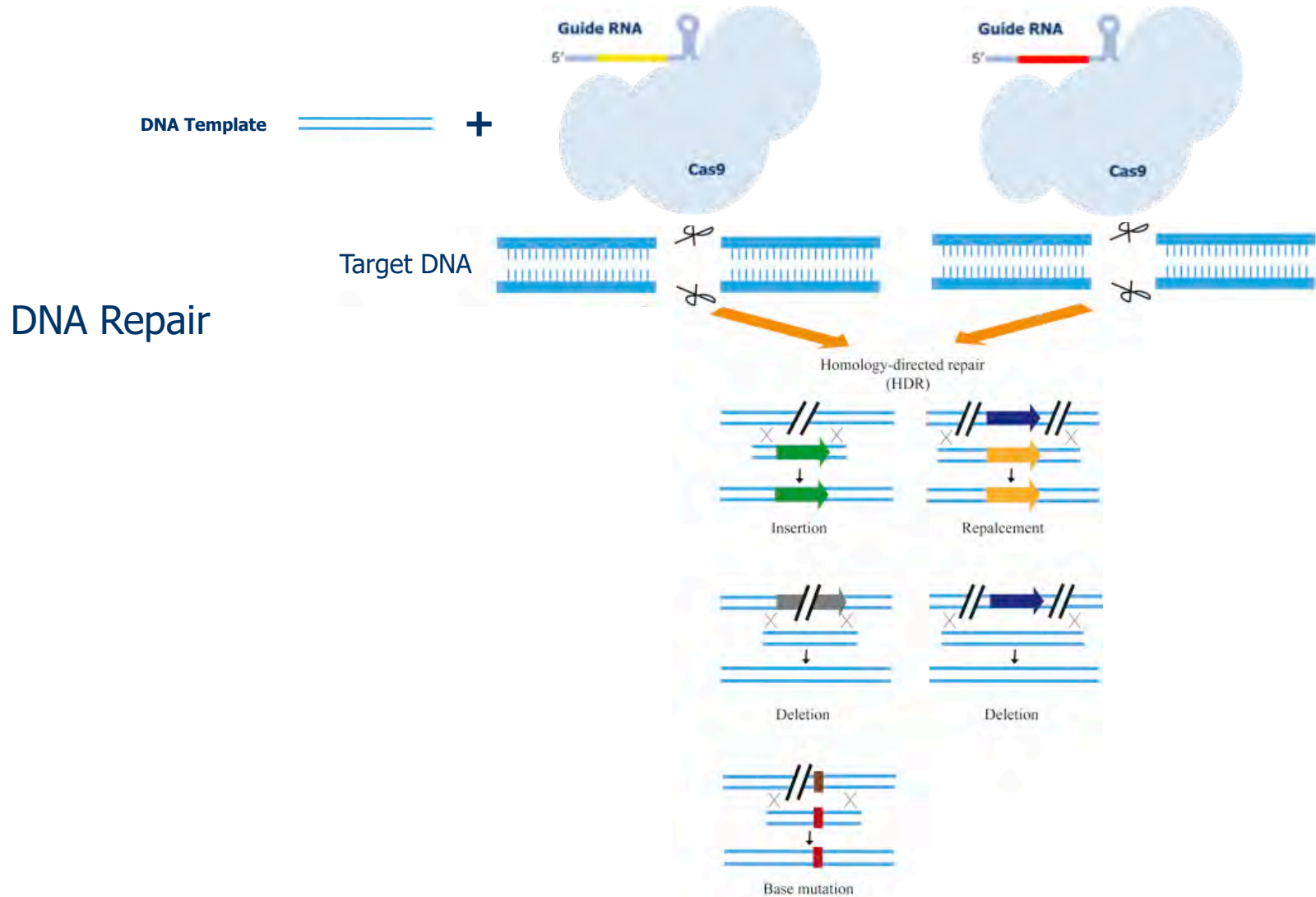
Francisco J.M. Mojica<sup>1,2</sup> and Lluís Montoliu<sup>3,4,\*</sup>

# Genome Editing

## DNA Repair



# Genome Editing



# Genome Editing

*Science*. 2013 February 15; 339(6121): 819–823. doi:10.1126/science.1231143.

## Multiplex Genome Engineering Using CRISPR/Cas Systems

Le Cong<sup>1,2,\*</sup>, F. Ann Ran<sup>1,4,\*</sup>, David Cox<sup>1,3</sup>, Shuailiang Lin<sup>1,5</sup>, Robert Barretto<sup>6</sup>, Naomi Habib<sup>1</sup>, Patrick D. Hsu<sup>1,4</sup>, Xuebing Wu<sup>7</sup>, Wenyan Jiang<sup>8</sup>, Luciano A. Marraffini<sup>8</sup>, and Feng Zhang<sup>1,†</sup>

*Science*. 2013 February 15; 339(6121): 823–826. doi:10.1126/science.1232033.

## RNA-Guided Human Genome Engineering via Cas9

Prashant Mali<sup>1,5</sup>, Luhan Yang<sup>1,3,5</sup>, Kevin M. Esvelt<sup>2</sup>, John Aach<sup>1</sup>, Marc Guell<sup>1</sup>, James E. DiCarlo<sup>4</sup>, Julie E. Norville<sup>1</sup>, and George M. Church<sup>1,2,\*</sup>



L. Marraffini

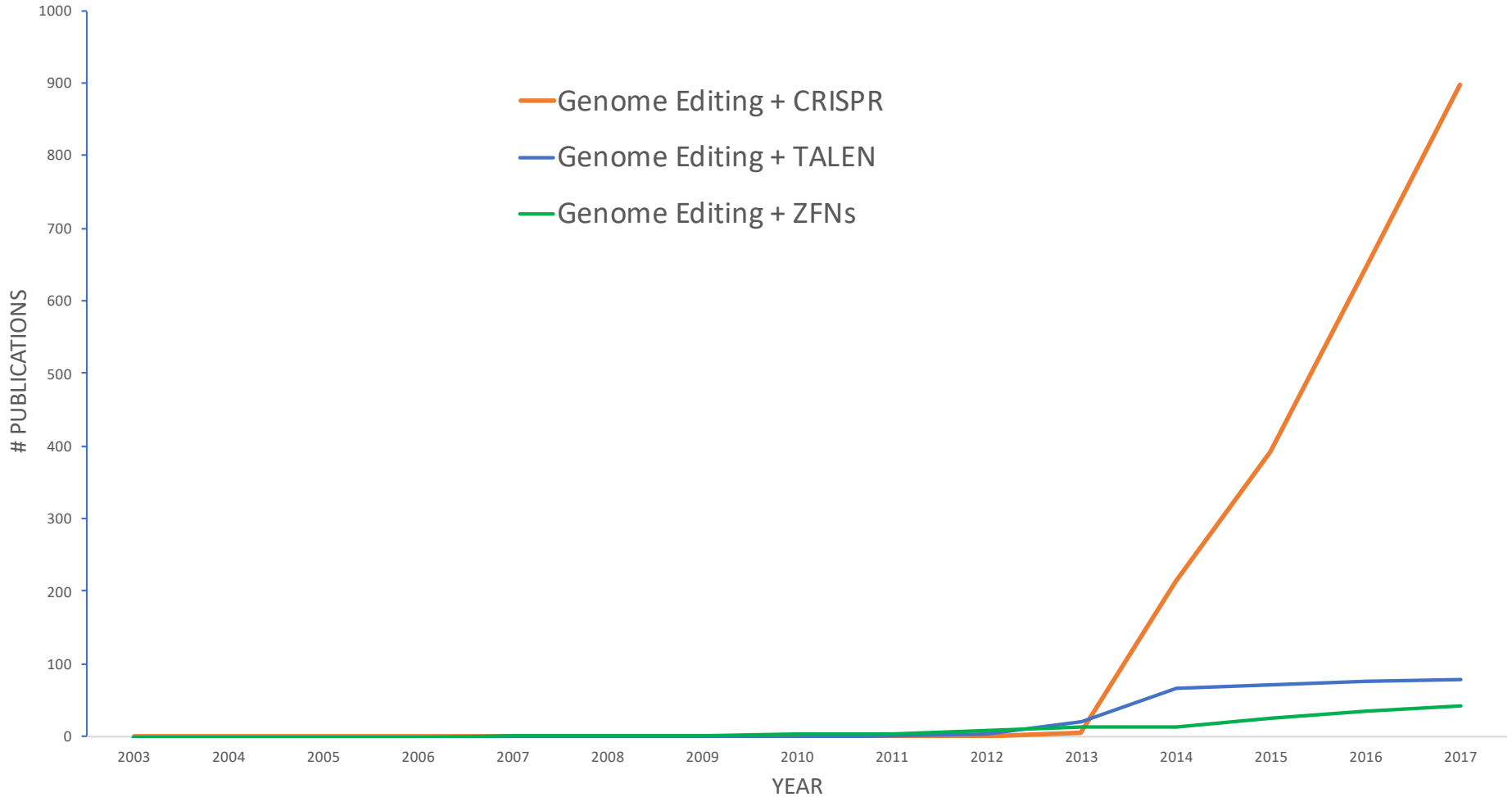


F. Zhang



G. Church

# TALENs , ZFN *et al.* Superseded by CRISPR





## Programmable editing of a target base in genomic DNA without double-stranded DNA cleavage

**Alexis C. Komor, Yongjoo B. Kim, Michael S. Packer, John A. Zuris & David R. Liu**

**[Affiliations](#) | [Contributions](#) | [Corresponding author](#)**

*Nature* (2016) | doi:10.1038/nature17946

Received 26 February 2016 | Accepted 30 March 2016 | Published online 20 April 2016

# “CRISPR-edited” organisms

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- **Bacteria**
- **Protozoa**: *Leishmania, Toxoplasma, Plasmodium, Trypanosoma...*
- **Fungi**: yeasts, molds, mushroom

- **Plants:**

Rice

Orange

Wheat

Soybean

Corn

Potato

Tomato

Tobacco

*Arabidopsis*

Grapefruit

Petunia

Cotton

Lettuce

Cucumber

Sorghum

# “CRISPR-edited” organisms

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•Worms

•Insects

•Reptiles

•Amphibians

•Molluscs

•Fish

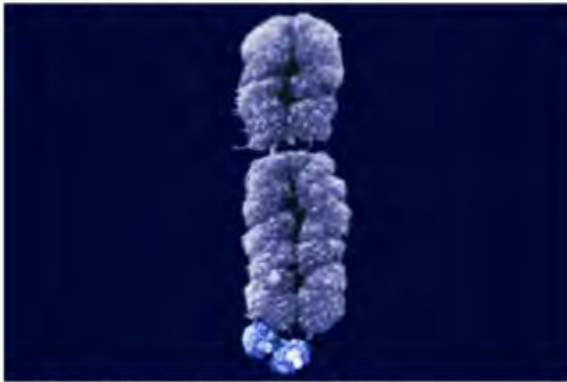
•Birds

•Mammals: mouse, rabbit, goat, rat, dog, pig, primate...

# Targeting Disease

DAILY NEWS 15 February 2018

**CRISPR has fixed the genetic cause  
of a learning disability**



Fragile X syndrome is the most common genetic cause of intellectual disability  
Christine Harrison/Visuals Unlimited, Inc./SCIENCE PHOTO LIBRARY

Cancer  
Malaria  
Albinism  
Cataracts  
Hemophilia  
Cystic fibrosis  
 $\beta$ -thalassemia  
Retinitis Pigmentosa  
Hypercholesterolemia  
Neurodegenerative disorders  
Duchenne Muscular Dystrophy  
Viral infections (HV, HBV, HPV, HIV...)


# Editing Human Embryos



Mol Genet Genomics (2017) 292:525–533  
DOI 10.1007/s00438-017-1299-z

ORIGINAL ARTICLE

## CRISPR/Cas9-mediated gene editing in human zygotes using Cas9 protein

Lichun Tang<sup>1,2</sup>  · Yanting Zeng<sup>3</sup> · Hongzi Du<sup>3</sup> · Mengmeng Gong<sup>1</sup> · Jin Peng<sup>1</sup> · Buxi Zhang<sup>1</sup> · Ming Lei<sup>3</sup> · Fang Zhao<sup>4</sup> · Weihua Wang<sup>5</sup> · Xiaowei Li<sup>6</sup> · Jianqiao Liu<sup>3</sup>

J Assist Reprod Genet (2016) 33:581–588  
DOI 10.1007/s10815-016-0710-8

TECHNOLOGICAL INNOVATIONS

## Introducing precise genetic modifications into human 3PN embryos by CRISPR/Cas-mediated genome editing

Xiangjin Kang<sup>1</sup> · Wenyin He<sup>1</sup> · Yuling Huang<sup>1</sup> · Qian Yu<sup>1</sup> · Yaoyong Chen<sup>1</sup> · Xingcheng Gao<sup>1</sup> · Xiaofang Sun<sup>1</sup> · Yong Fan<sup>1</sup>

## ARTICLE

doi:10.1038/nature24033

## Genome editing reveals a role for OCT4 in human embryogenesis

Norah M. E. Fogarty<sup>1</sup>, Afshan McCarthy<sup>1</sup>, Kirsten E. Snijders<sup>2</sup>, Benjamin E. Powell<sup>1</sup>, Nada Kubikova<sup>4</sup>, Paul Blakeley<sup>1</sup>, Rebecca Lea<sup>3</sup>, Kay Elder<sup>3</sup>, Sissy E. Wamaitha<sup>1</sup>, Daesik Kim<sup>6</sup>, Valdone Maciulyte<sup>3</sup>, Jens Kleinjung<sup>7</sup>, Jin-Soo Kim<sup>8,9</sup>, Dagan Wells<sup>4</sup>, Ludovic Vallier<sup>2,9,10</sup>, Alessandro Bertero<sup>10</sup>†, James M. A. Turner<sup>3</sup> & Kathy K. Niakan<sup>1</sup>

## ARTICLE

doi:10.1038/nature23305

## Correction of a pathogenic gene mutation in human embryos

Hong Ma<sup>1\*</sup>, Nuria Marti-Gutierrez<sup>2\*</sup>, Sang-Wook Park<sup>2\*</sup>, Jun Wu<sup>3\*</sup>, Yeonmi Lee<sup>1</sup>, Keiichiro Suzuki<sup>3</sup>, Amy Koski<sup>1</sup>, Dongmei Ji<sup>1</sup>, Tomonari Hayama<sup>1</sup>, Riffat Ahmed<sup>1</sup>, Hayley Darby<sup>1</sup>, Crystal Van Dyken<sup>1</sup>, Ying Li<sup>1</sup>, Eunju Kang<sup>1</sup>, A.-Reum Park<sup>2</sup>, Daesik Kim<sup>4</sup>, Sang-Tae Kim<sup>2</sup>, Jianhui Gong<sup>5,6,7,8</sup>, Ying Gu<sup>5,6,7</sup>, Xun Xu<sup>5,6,7</sup>, David Battaglia<sup>1,9</sup>, Sacha A. Krieg<sup>9</sup>, David M. Lee<sup>9</sup>, Diana H. Wu<sup>9</sup>, Don P. Wolf<sup>1</sup>, Stephen B. Heitner<sup>10</sup>, Juan Carlos Izpisua Belmonte<sup>3</sup>§, Paula Amato<sup>1,9</sup>§, Jin-Soo Kim<sup>2,4</sup>§, Sanjiv Kaul<sup>10</sup>§ & Shoukhrat Mitalipov<sup>1,10</sup>§

Gene Ther. 2016 May 19. doi: 10.1038/gt.2016.41. [Epub ahead of print]

## Excision of HIV-1 DNA by gene editing: a proof-of-concept in vivo study.

Kaminski R<sup>1</sup>, Bella R<sup>2</sup>, Yin C<sup>1</sup>, Otte J<sup>1</sup>, Ferrante P<sup>2</sup>, Gendelman HE<sup>3</sup>, Li H<sup>4</sup>, Booze R<sup>4</sup>, Gordon J<sup>1</sup>, Hu W<sup>1</sup>, Khalili K<sup>1</sup>.

## LETTER

doi:10.1038/nature25164

### Treatment of autosomal dominant hearing loss by in vivo delivery of genome editing agents

Xue Gao<sup>1,2,3,\*</sup>, Yong Tao<sup>4,5,\*</sup>, Veronica Lamas<sup>4</sup>, Mingqian Huang<sup>4</sup>, Wei-Hsi Yeh<sup>1,2,3,6</sup>, Bifeng Pan<sup>7</sup>, Yu-Juan Hu<sup>4,5</sup>, Johnny H. Hu<sup>1,2,3</sup>, David B. Thompson<sup>1,2</sup>, Yilai Shu<sup>4,8</sup>, Yamin Li<sup>9</sup>, Hongyang Wang<sup>4,10</sup>, Shiming Yang<sup>10</sup>, Qiaobing Xu<sup>9</sup>, Daniel B. Polley<sup>4</sup>, M. Charles Liberman<sup>4</sup>, Wei-Jia Kong<sup>6</sup>, Jeffrey R. Holt<sup>7</sup>, Zheng-Yi Chen<sup>6,8</sup> & David R. Liu<sup>1,2,3</sup>

Sci Adv. 2017 Dec 20;3(12):eaar3952. doi: 10.1126/sciadv.aar3952. eCollection 2017 Dec.

### In vivo genome editing improves motor function and extends survival in a mouse model of ALS.

Gaj T<sup>1</sup>, Qjala DS<sup>2</sup>, Ekman FK<sup>3</sup>, Byrne LC<sup>4</sup>, Limsirichai P<sup>5</sup>, Schaffer DV<sup>1,2,4</sup>.

## ScienceDaily

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### Science News

from research organizations

### New gene-editing technology partially restores vision in blind animals

Date: November 16, 2016

Source: Salk Institute

**Summary:** Researchers have discovered a holy grail of gene editing -- the ability to, for the first time, insert DNA at a target location into the non-dividing cells that make up the majority of adult organs and tissues. The technique, which the team showed was able to partially restore visual responses in blind rodents, will open new avenues for basic research and a variety of treatments, such as for retinal, heart and neurological diseases.

### Research Article



EMBO  
Molecular Medicine

### CRISPR/Cas9-mediated somatic correction of a novel coagulator factor IX gene mutation ameliorates hemophilia in mouse

Yuting Guan<sup>1,†</sup>, Yanlin Ma<sup>2,3,†,\*</sup>, Qi Li<sup>2</sup>, Zhenliang Sun<sup>4</sup>, Lie Ma<sup>1</sup>, Lijuan Wu<sup>1</sup>, Liren Wang<sup>1</sup>, Li Zeng<sup>1</sup>, Yanjiao Shao<sup>1</sup>, Yuting Chen<sup>1</sup>, Yu<sup>3</sup>.

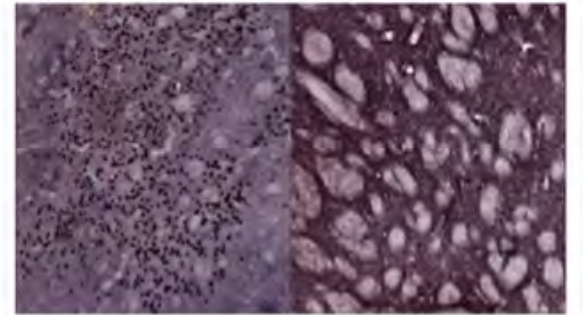


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News > Biology > Gene-editing method halts production of brain-destroying proteins

### LATEST NEWS

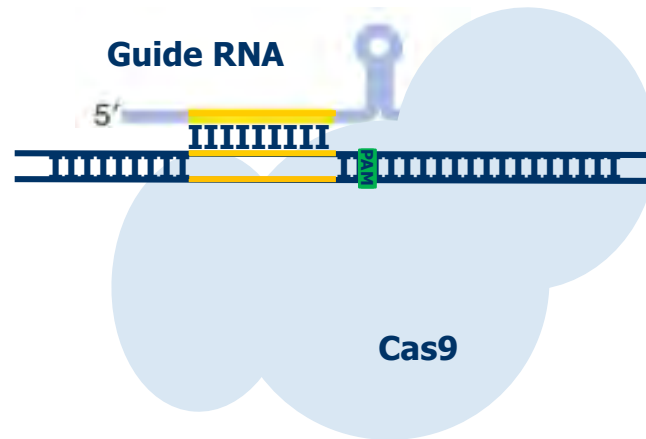


The brain of an untreated mouse on the left, showing huntingtin protein aggregation (a hallmark of Huntington's disease) and on the right, the brain of a mouse treated with CRISPR-Cas9 editing, showing the lack of protein aggregation.

### Gene-editing method halts production of brain-destroying proteins

# Clinical Trials with CRISPR

Row	Saved	Status	Study Title	Conditions	Interventions
1		Not yet recruiting	<a href="#">A Safety and Efficacy Study of TALEN and CRISPR/Cas9 in the Treatment of HPV-related Cervical Intraepithelial Neoplasia</a>	<ul style="list-style-type: none"> <li>Human Papillomavirus-Related Malignant Neoplasm</li> </ul>	<ul style="list-style-type: none"> <li>Biological: TALEN</li> <li>Biological: <b>CRISPR/Cas9</b></li> </ul>
2		Recruiting	<a href="#">Safety of Transplantation of CRISPR CCR5 Modified CD34+ Cells in HIV-infected Subjects With Hematological Malignances</a>	<ul style="list-style-type: none"> <li>HIV-1-infection</li> </ul>	<ul style="list-style-type: none"> <li>Genetic: CCR5 gene modification</li> </ul>
3		Not yet recruiting	<a href="#">Examining the Knowledge, Attitudes, and Beliefs of Sickle Cell Disease Patients, Parents of Patients With Sickle Cell Disease, and Providers Towards the Integration of CRISPR in Clinical Care</a>	<ul style="list-style-type: none"> <li>Sickle Cell Disease</li> </ul>	
4		Recruiting	<a href="#">NY-ESO-1-redirected CRISPR (TCRendo and PD1) Edited T Cells (NYCE T Cells)</a>	<ul style="list-style-type: none"> <li>Multiple Myeloma</li> <li>Melanoma</li> <li>Synovial Sarcoma</li> <li>Myxoid/Round Cell Liposarcoma</li> </ul>	<ul style="list-style-type: none"> <li>Biological: NY-ESO-1 redirected autologous T cells with <b>CRISPR</b> edited endogenous TCR and PD-1</li> <li>Drug: Cyclophosphamide</li> <li>Drug: Fludarabine</li> <li>Device: NY-ESO-1 expression testing</li> </ul>
5		Recruiting	<a href="#">Identification of Host Factors of Norovirus Infections in Mini-Gut Model</a>	<ul style="list-style-type: none"> <li>Gastrointestinal Infection</li> </ul>	<ul style="list-style-type: none"> <li>Procedure: Duodenal biopsy</li> <li>Procedure: Saliva</li> </ul>
6		Recruiting	<a href="#">A Study Evaluating UCART019 in Patients With Relapsed or Refractory CD19+ Leukemia and Lymphoma</a>	<ul style="list-style-type: none"> <li>B Cell Leukemia</li> <li>B Cell Lymphoma</li> </ul>	<ul style="list-style-type: none"> <li>Biological: UCART019</li> </ul>
7		Recruiting	<a href="#">A Feasibility and Safety Study of Universal Dual Specificity CD19 and CD20 or CD22 CAR-T Cell Immunotherapy for Relapsed or Refractory Leukemia and Lymphoma</a>	<ul style="list-style-type: none"> <li>B Cell Leukemia</li> <li>B Cell Lymphoma</li> </ul>	<ul style="list-style-type: none"> <li>Biological: Universal Dual Specificity CD19 and CD20 or CD22 CAR-T Cells</li> </ul>
8		Recruiting	<a href="#">PD-1 Knockout Engineered T Cells for Advanced Esophageal Cancer</a>	<ul style="list-style-type: none"> <li>Esophageal Cancer</li> </ul>	<ul style="list-style-type: none"> <li>Other: PD-1 Knockout T Cells</li> </ul>



## No PAM

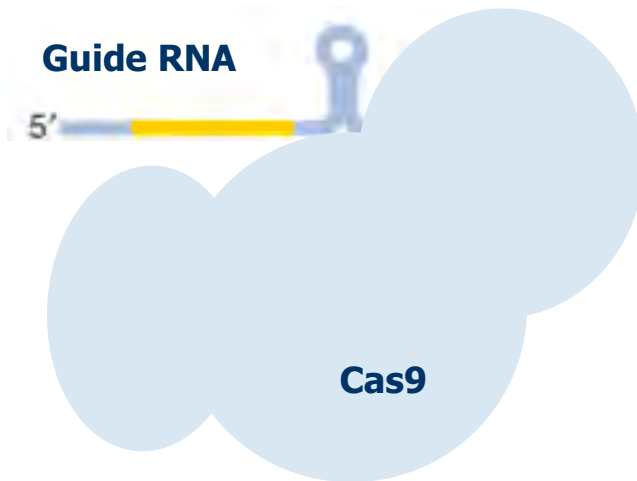
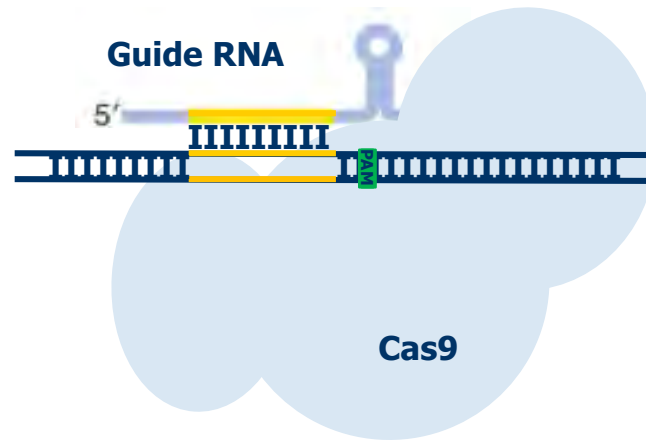


Table 2  
Reported Cas9 variants and orthologues

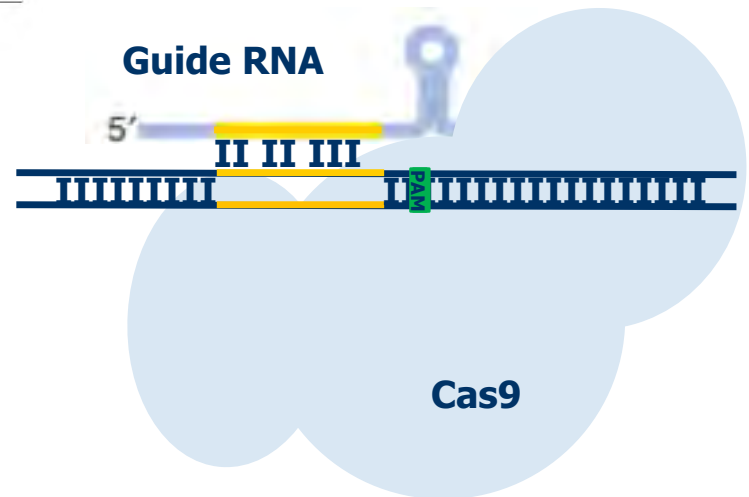
Name	Origin	Primary PAM
SpCas9	<i>Streptococcus pyogenes</i>	NGG (3' to protospacer) <sup>a</sup>
SpCas9-VQR	Engineered	NGAG (3' to protospacer)
SpCas9-VRER	Engineered	NGCG (3' to protospacer)
eSpCas9 (1.1)	Engineered	NGG (3' to protospacer)
NmCas9	<i>Neisseria meningitidis</i>	NNNGATT (3' to protospacer)
St1Cas9	<i>Streptococcus thermophiles</i>	NNAGAA (3' to protospacer)
SaCas9	<i>Staphylococcus aureus</i>	NNGGGT (3' to protospacer)
AsCpf1	<i>Acidaminococcus sp. BV3L6</i>	TTTN (5' to protospacer)
BICas9	<i>Brevibacillus laterosporus</i>	NNNNCNA (3' to protospacer)

Zhang, D., et al., Journal of Genetics and Genomics (2016)





## Off-Targets

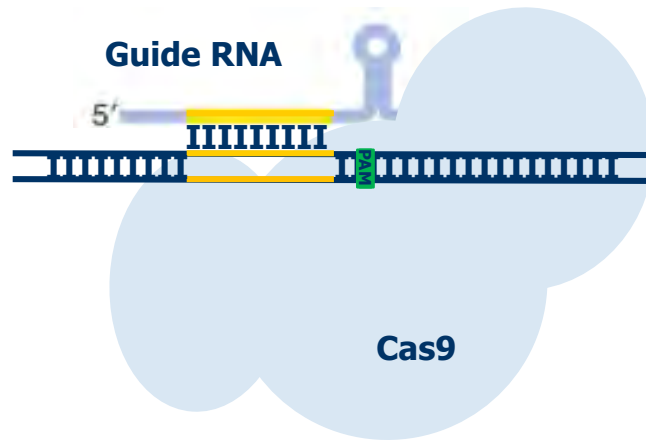


## ARTICLE

doi:10.1038/nature16526

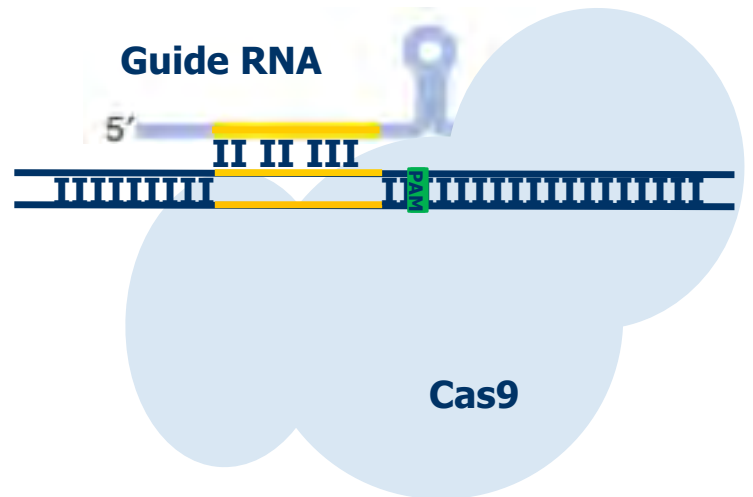
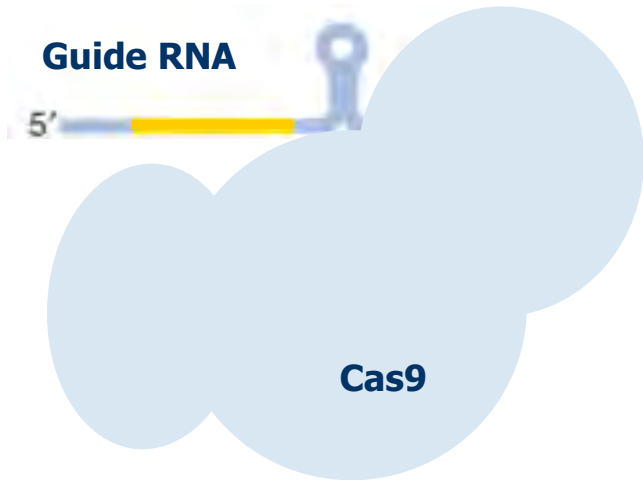
# High-fidelity CRISPR-Cas9 nucleases with no detectable genome-wide off-target effects

Benjamin P. Kleinstiver<sup>1,2\*</sup>, Vikram Pattanayak<sup>1,2\*</sup>, Michelle S. Prew<sup>1</sup>, Shengdar Q. Tsai<sup>1,2</sup>, Nhu T. Nguyen<sup>1</sup>, Zongli Zheng<sup>3</sup> & J. Keith Joung<sup>1,2</sup>



**No PAM**

**Off-Targets**



# ARTICLE

doi:10.1038/nature26155

## Evolved Cas9 variants with broad PAM compatibility and high DNA specificity

Johnny H. Hu<sup>1,2,3</sup>, Shannon M. Miller<sup>1,2,3</sup>, Maarten H. Geurts<sup>1,2,3</sup>, Weixin Tang<sup>1,2,3</sup>, Liwei Chen<sup>1,2,3</sup>, Ning Sun<sup>1,2,3</sup>, Christina M. Zeina<sup>1,2,3</sup>, Xue Gao<sup>1,2,3</sup>, Holly A. Rees<sup>1,2,3</sup>, Zhi Lin<sup>1,2,3</sup> & David R. Liu<sup>1,2,3</sup>

# On-Target Variations

	sgRNA 5'0	sgRNA 5'5
reference:	TCCCAAGAATTAAGTGTGACAGTGCAAGAT***TTT <b>TGG</b> CCTAGAGCTTTGCAGTGATGATTA	
TYRINS5#19_1:	TCCCAAGAATTAAGTGTGACAG-----	(+120-8) -- <b>TGG</b> CCTAGAGCTTTGCAGTGATGATTA
TYRINS5#19_2:	TCCCAAGAATTAAGTGTGACAG-----	(+120-8) -- <b>TGG</b> CCTAGAGCTTTGCAGTGATGATTA
TYRINS5#23_1:	TCCCAAGAATTAAGTGTGACAGTGCAAGA	(+6-2) <b>TTTGG</b> CCTAGAGCTTTGCAGTGATGATTA
TYRINS5#23_2:	TCCCAAGAATTAAGTGTGACAGTGCAAGA	(+6-2) <b>TTTGG</b> CCTAGAGCTTTGCAGTGATGATTA
TYRINS5#27_1:	TCCCAAGAATTAAGTGTGACAGTGCAAGAT***	<b>TTTGG</b> CCTAGAGCTTTGCAGTGATGATTA
TYRINS5#27_2:	TCCCAAGAATTAAGTGTGACAGTGCAAGAT***	<b>TTTGG</b> CCTAGAGCTTTGCAGTGATGATTA
TYRINS5#30_1:	TCCCAAGAATTAAGTGTGACAGTGCAAGAT(+5)	<b>TTTGG</b> CCTAGAGCTTTGCAGTGATGATTA
TYRINS5#30_2:	TCCCAAGAATTAAGTGTGACAGTGCAAGAT(+5)	<b>TTTGG</b> CCTAGAGCTTTGCAGTGATGATTA
TYRINS5#31_3:	TCCCAAGAATTAAGTGTGACAGTGCAAGA-***	-----GCTTTGCAGTGATGATTA
TYRINS5#41_1:	TCCCAAGAATTAAGTGTGAT <b>A</b> -----	*** <b>TTTGG</b> CCTAGAGCTTTGCAGTGATGATTA
TYRINS5#41_1:	TCCCAAGAATTAAGTGTGAT <b>A</b> -----	*** <b>TTTGG</b> CCTAGAGCTTTGCAGTGATGATTA

Cite as: D. B. T. Cox *et al.*, *Science*  
10.1126/science.aaq0180 (2017).

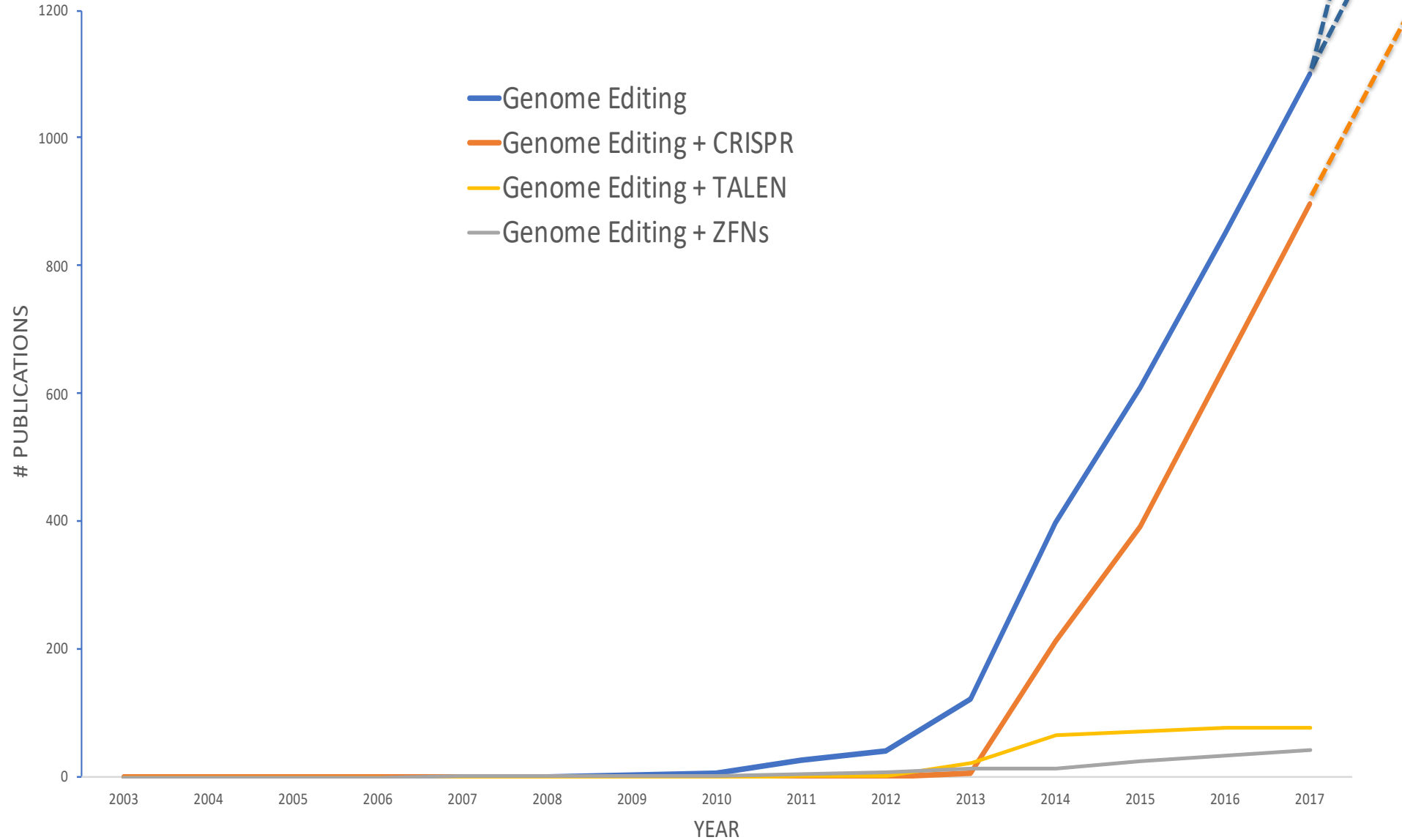
# RNA editing with CRISPR-Cas13

David B. T. Cox,<sup>1,2,3,4,5,6\*</sup> Jonathan S. Gootenberg,<sup>1,2,3,4,7\*</sup> Omar O. Abudayyeh,<sup>1,2,3,4,6\*</sup> Brian Franklin,<sup>1,2,3,4</sup> Max J. Kellner,<sup>1,2,3,4</sup> Julia  
Joung,<sup>1,2,3,4</sup> Feng Zhang<sup>1,2,3,4†</sup>



**Feng Zhang**

# Genome Editing Papers in PubMed



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10.1126/science.aar4120 (2018).

# Systematic discovery of antiphage defense systems in the microbial pangenome

Shany Doron,\* Sarah Melamed,\* Gal Ofir, Azita Leavitt, Anna Lopatina, Mai Keren, Gil Amitai, Rotem Sorek<sup>†</sup>