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# Genome editing in laboratory animals, ethical use and welfare challenge.

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Mary Lyon Centre – MRC Harwell Institute

**ARRIGE meeting, Paris, 14/11/19**

# The Mary Lyon Centre – MRC Harwell Institute



- Over 20 years of experience in genome engineering
- Generation of new mutants (>150 new models/year)
- Breeding (>50,000 mice housed)
- Archiving and distribution of mice
- I am a user (and developer) of genome editing, not a philosopher. There are more questions than answers.



Premises of this talk:

1. We are using research on animals to improve human health

2. We ought to use research on animals to improve human health



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- Genome Editing, a disruptive technology
  - 3Rs
  - The 4<sup>th</sup> R
  - The conversation



# The special case of Genome Editing

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- How is genome editing a disruptive technology?
- How does genome editing change how we implement the ethical imperatives of the 3Rs?



- **The bespoke (mouse) model of a rare disease:**

- > patient perspective (“their mutation”, Acceptable/Wished for that research is undertaken ‘In their name’)? Counselling?
- > when is sufficient evidence obtained to justify the creation of an animal model?

- **The increased sophistication of the animal model and the animal house**



- **The mouse as a pre-clinical research tool for exploring the safety and efficacy of human assisted reproductive technologies**



-> *The new genome editing technologies trigger new ethical debates for our societies.*



# The special case of Genome Editing

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- How is genome editing a disruptive technology?
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- **Reduction: Distinguishing between per-experiment reduction and overall reduction**

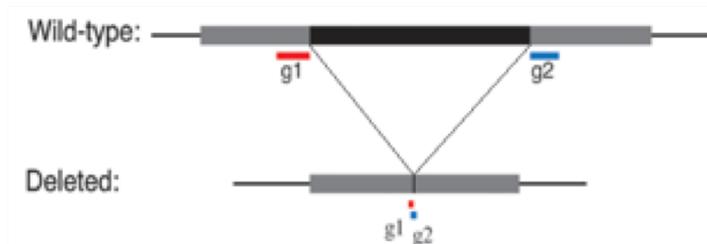
- **Indel(s) / Knock-out(s)** ✓



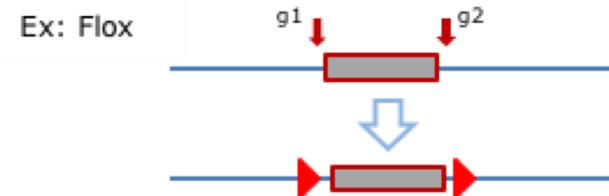
- **Point mutation(s)** ✓



- **Tailored deletion(s)** ✓



- **Gene targeting** ✓





# Genome Editing capacity: 150 new lines/year



- Over 250 projects in the pipeline, 180 analysed:

	<b>Indel</b>	<b>Deletion</b>	<b>PM</b>
Percentage of mutated founders / F0 born	<b>38.5</b>	<b>ND</b>	<b>24.6</b>
Percentage of expected mutants / F0 born	<b>28.5</b>	<b>15.7</b>	<b>6.8</b>
Frequency of animal with desired mutation	<b>1/3.5</b>	<b>1/6</b>	<b>1/15</b>
Completed projects so far	<b>22</b>	<b>56</b>	<b>32</b>
Positive founder produced	<b>8</b>	<b>31</b>	<b>14</b>

- **Refinement: Know your model's limitations.**

Example of phenotyping of  $G_0$ s

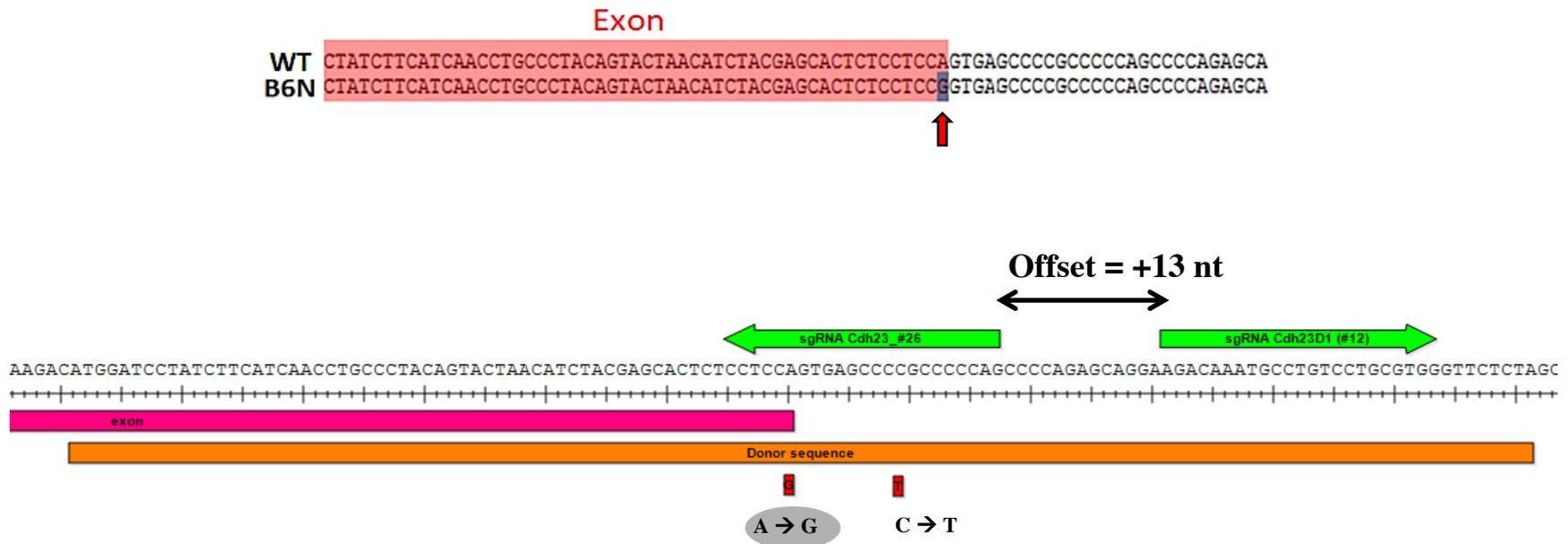
“the CRISPR/Cas system allows the **one-step generation** of animals carrying mutations in multiple genes, an approach that will greatly accelerate the in vivo study of functionally redundant genes and of epistatic gene interactions.”  
(Wang et al. 2013, *Cell*).



# Generation of a point mutation with CRISPR

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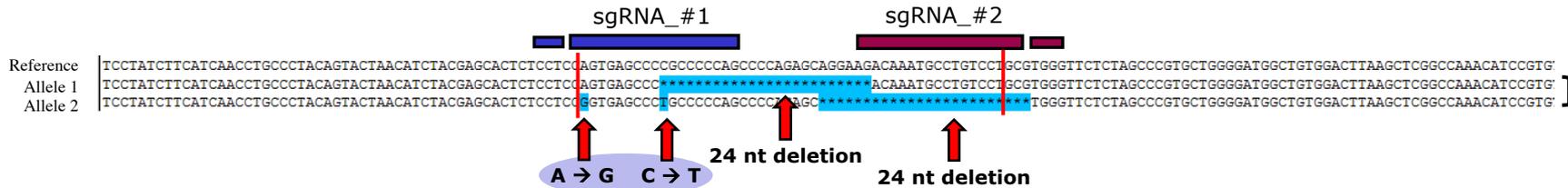
Mutation repair of *Cdh23*<sup>753A</sup> SNP associated with Age-related Hearing Loss (AHL)





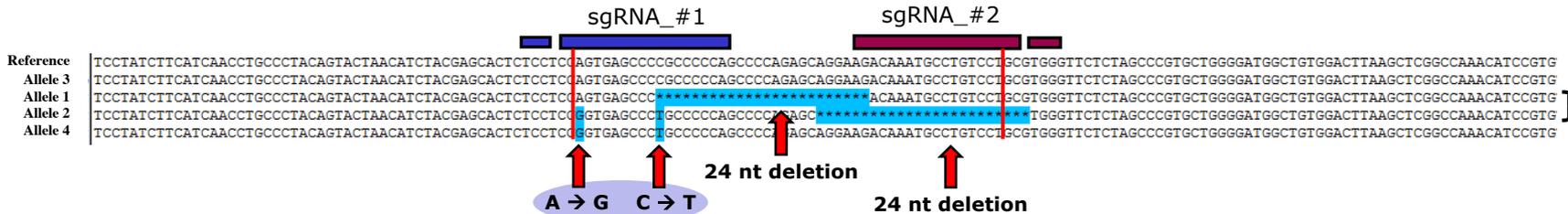
# Unpredictable repair and mosaicism in G<sub>0</sub> mice

## Cdh23 target, F0 #30 ear clip genotyping results:



- Allele 1 = NHEJ repair → 24 nt deletion
- Allele 2 = Illegitimate repair → Correct repair at the target + 24 nt deletion

## Cdh23 target, alleles found in F1s (#30 offspring):



- Allele 1 = NHEJ repair → 24 nt deletion
- Allele 2 = Illegitimate repair → Correct repair at the target + 24 nt deletion
- Allele 3 = WT
- Allele 4 = HDR → Correctly repaired



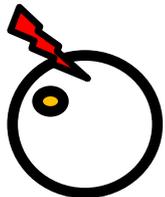
# Genotype complexity and mosaicism

- Cas9 mRNA
- sgRNA(s)
- DNA donor template

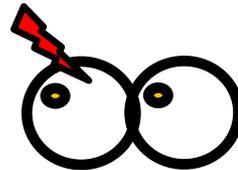
Pronuclear injection in 1 cell stage embryos



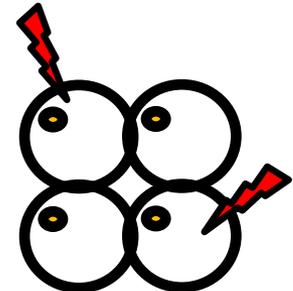
 = Mutagenesis event



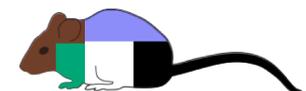
Mutant heterozygous  
(2 alleles)



Mutant mosaic  
(3 alleles)

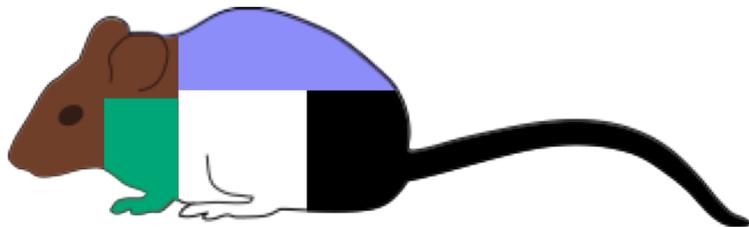


Mutant mosaic  
(5 alleles)





# Founder animals can display welfare issue





- **Replacement:**

Increased accessibility of other models, including human and other species – places a greater onus on scientists to justify use of a particular animal model.

A better model could be a more evolved organism.



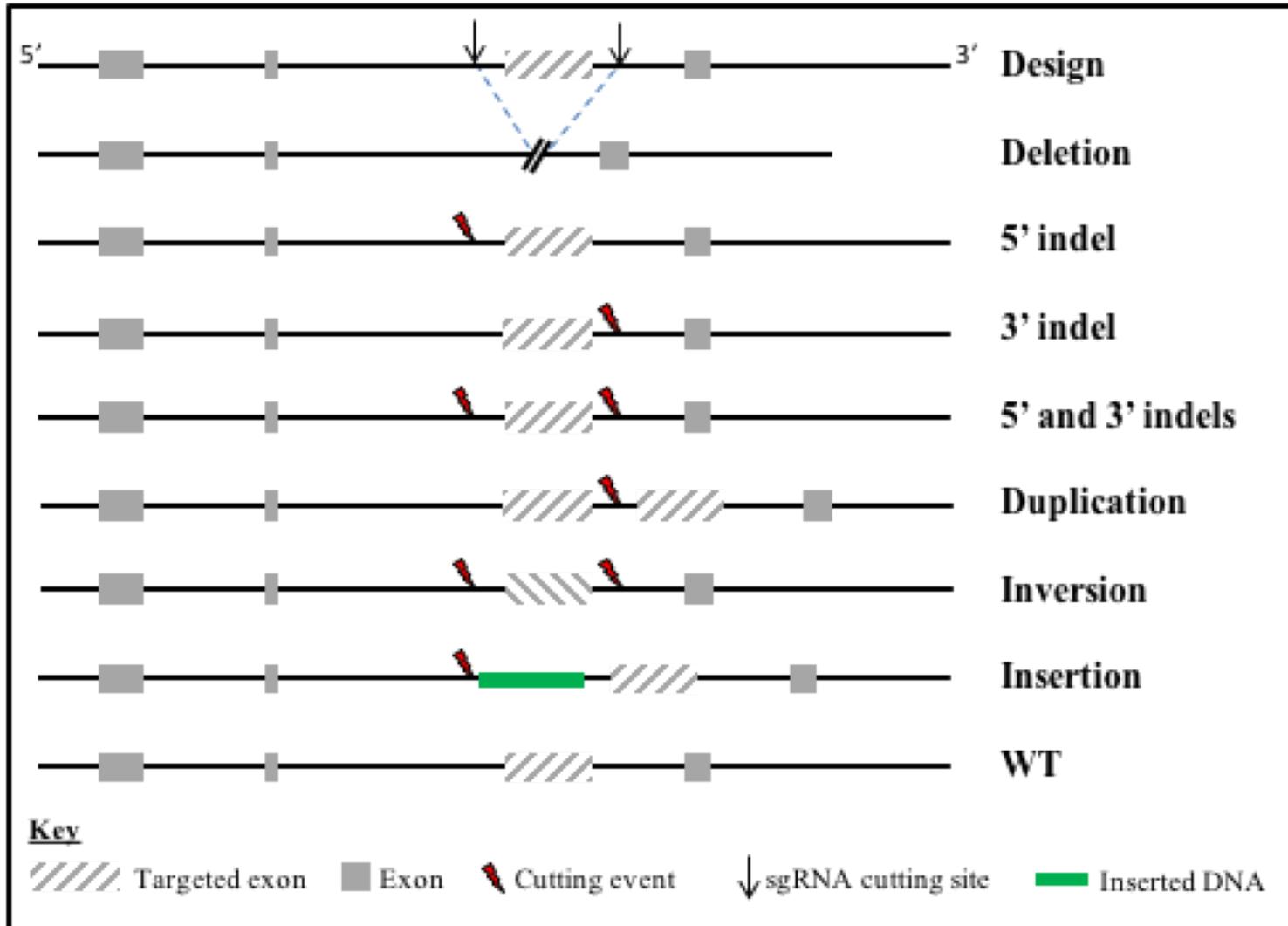


- **Reproducibility:**

There can not be any justification for the use of animals in research if the research is not reproducible.

**Genome editing tools just cut DNA, they do not control the sequence of the new mutations.**

# Use of 2 sgRNAs can yield many artefacts

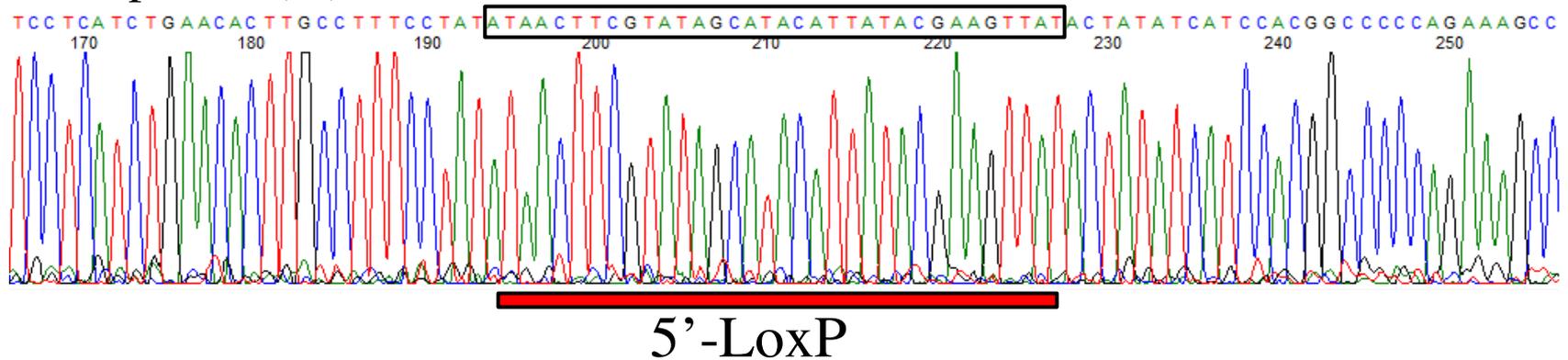




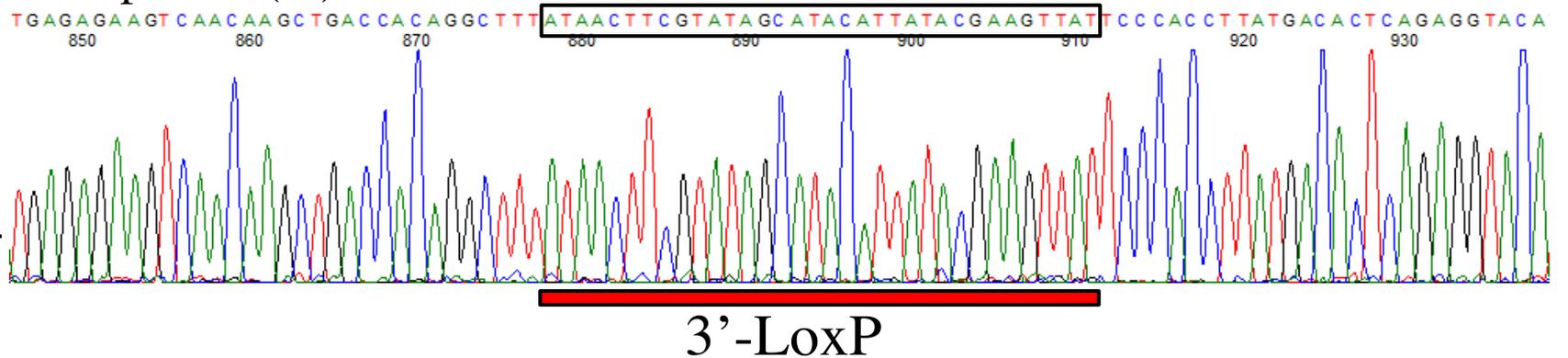
# Conditional allele: Full validation is a complex exercise

- $G_0$  #2  $\rightarrow$  Homozygous for the repair?

Forward primer (5')



Reverse primer (3')



Critical region all correct!

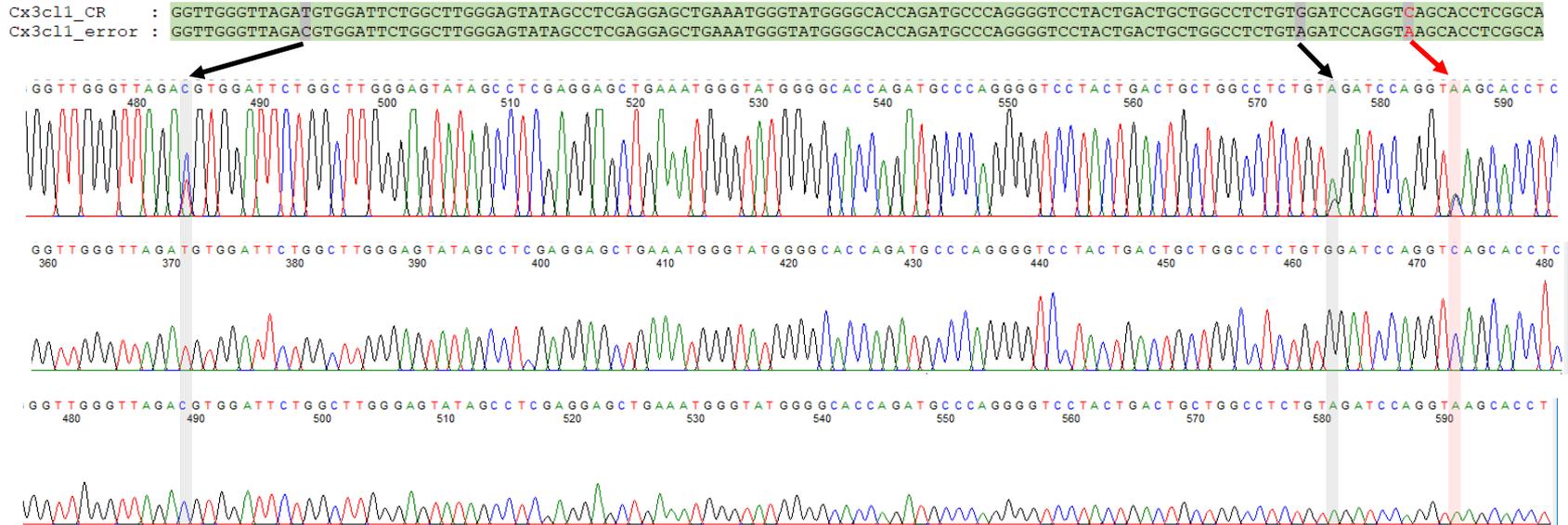


# Beware of additional sequence changes

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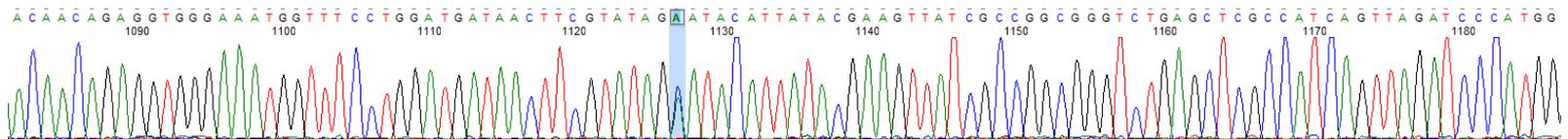
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b



c

Cx3cl1\_flox : ACAACAGAGGTGGGAAATGGTTTCCTGGATGATAACTTCGTATAGCATACATTATACGAAGTTATCGCCGGCGGGTCTGAGCTCGCCATCAGTTAGATCCCATGG  
Cx3cl1\_error : ACAACAGAGGTGGGAAATGGTTTCCTGGATGATAACTTCGTATAGATAACATTATACGAAGTTATCGCCGGCGGGTCTGAGCTCGCCATCAGTTAGATCCCATGG





# Analysis of a new cKO line

Animal	PCR Result	ddPCR result
F <sub>0</sub> #2	Correct mutant: Homozygous	2.78 (3 copies)



Breeding gave rise to 8 F1 mice

Animal	PCR Result	ddPCR result
#2 offspring 1	WT	1
#2 offspring 2	WT	2
#2 offspring 3	Correct mutant	3
#2 offspring 4	WT	3
#2 offspring 5	Correct mutant	2
#2 offspring 6	Correct mutant	2
#2 offspring 7	Correct mutant	3
#2 offspring 8	WT	3

Evidence of additional random integration of the donor.



# Genome editing versatility

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	<i>Syt7</i>	<i>Ikzf2</i>	<i>USP45</i>	<i>Syt4</i>	<i>Rapgef5</i>
ssDNA size (nt)	1149	1107	914	1567	899
Nb of pups	17	17	19	67	30
Nb of mutants	10	5	6	32	12
Mutant with exon deletion	3	3	2	3	5
Animals with 2 sides cut	29%	24%	11%	9%	20%
<b>Mutant with floxed allele</b>	<b>2</b>	<b>1</b>	<b>0</b>	<b>3</b>	<b>1</b>
GLT	Yes	Yes		Yes	Founder dead

= a greater burden on allele validation



# Standards for allele validation

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- H  $G_0$ s should be treated as mosaic and can be screened for the presence of a mutation.
- H New alleles should be fully sequenced in  $G_1$ s.
- H Model validation should also include
  - copy counting of deleted segments,
  - copy counting of donor sequences,
  - Sequencing of off-target sites linked to locus of interest.



The challenge of genome editing is becoming **validation**.



## **Society**

- Importance of engaging stakeholders and “public”

## **Professionals**

- Researchers, medics and animal technologists



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- Important because political legitimisation is essential.
  - Does involve offering an ethical justification.
  - Requires a lot of engagement from scientists on this issue.
  - Literature on the ethics of animal experimentation (speciesism, utilitarianism, animal rights etc) all still applies.
  - Acknowledge the technology underpins working in close relation with the clinic (and therefore the patients!).



# Importance of training of researchers, medics and animal technologists

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- Expertise: This is still a new technology, we are all learning about it.
- Need to agree on standard for best practice: ie case of archiving, how do we control genetic integrity of new models.
- Not all about regulation, character and culture (Greenfield, 2017, Mammalian Genome).



“We acknowledge that the UK has the most detailed legislative framework regarding animal research in the world. But...regulation can act as an emotional screen between the researcher and an animal, possibly encouraging researchers to believe that *simply to conform to regulations is to act in a moral way*. It is therefore crucial to promote best practice more actively and to **improve the culture of care** in establishments licensed to conduct experiments using animals.”

NCOB, 2005 – “The Ethics of Research Involving Animals”



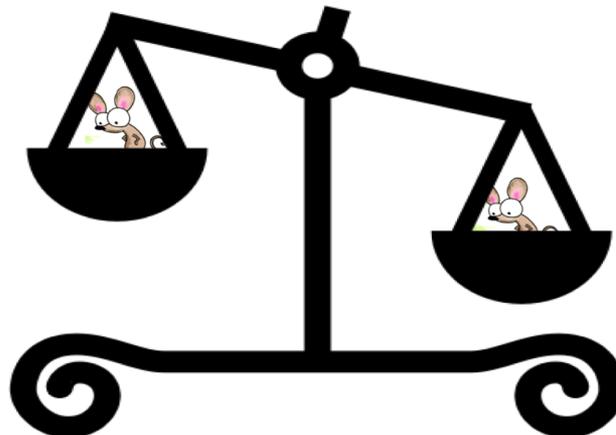
- New technology brings new questions: ethics, 3Rs application, but also thinking of research reproducibility.
- Facing important debates both in society and in scientific community.

## Cost

Animal suffering

Research Resources

Public Opinion



## Benefits

Health

Fundamental  
Knowledge

Engagement in  
Research



Many thanks!



## **MCB group at Mary Lyon Centre**

Gemma Codner

Joffrey Mianné

## **Colleagues in genome engineering field**

## **Andy Greenfield (Harwell)**



The Mary Lyon Centre

